Functionalized Inorganic Nanoparticles for the Detection of Food and Waterborne Bacterial Pathogens

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Abstract Infections acquired from ingesting contaminated food and water poses an adverse effect on public health and safety, thus affecting nations’ economy. Technical approaches developed over years have contributed adequately to microbial detection in food and water, yet, unveiling spaces for more improvement on early and rapid detection of pathogens. This review highlights different strategy assessing bio-functionalized inorganic nanoparticles towards the detection of pathogens in food and water samples. Conjugates of several bio-receptors and inorganic nanoparticles showed rapid, real-time, repeatability, and appreciable limit of detection in targeted pathogens. A patent referenced in this study established the biocompatibility of bio-functionalized inorganic nanoparticles mechanism. Unique attributes exhibited by bio-functionalized inorganic nanoparticles showed potential and improvement of the existing bio-sensing pathogen detection methods. Each of the identified strategies described showed a promising pathway accommodating the development of simple, and even the fabrication of low-cost materials for easy detection of bacterial pathogens in food and water products.

Keywords: foodborne infections, waterborne infections, bacterial detection, functionalized inorganic nanoparticles, Bioreceptors


1. Food and Waterborne Pathogen and Its Effect

Pathogenic bacteria from time immemorial and presently are still of significant concern to human health and safety, as they cause deleterious changes to man healthy living [1]. Although they are ubiquitous, their presence in food and water poses more harmful health risk resulting in both mild and fatal diseases even at a low infectious dose [2]. Some common illnesses and leading causal agents associated with food and water contamination globally are acute gastroenteritis, food poisoning (Staphylococcus aureus, Clostridium perfringens, Bacillus cereus), Botulism (Clostridium botulinum), Campylobacteriosis (Campylobacter jejuni), Listeriosis (Listeria monocytogenes), Salmonellosis (Salmonella spp), Hemorrhagic colitis (Escherichia coli O157:H7), Cholera (Vibrio cholera), diarrhoea (Vibrio parahaemolyticus) among others [3,4,5].

Among the pathogens identified as causal agents in food- and waterborne illnesses, enteric pathogens namely; Salmonella spp, Shigella spp, Yersinia enterolitica, Aeromonas spp and pathogenic Escherichia coli have been incriminated as the major pathogens causing hospitalizations, and even deaths due to the production of toxins and other cell metabolites within their host [6,7]. In addition to the severity of infections experienced by humans, the financial burden of food and water-borne infections has had some negative impact on individual’s income and nation’s economy. Areas affected majorly include: increase in medical expenses, productivity loss and loss of human resources [8]. Its financial implication is estimated to cause a loss of about $15.5 billion in the US, and $110.2 billion in low- and middle-income countries each year [8,9]. The impact of waterborne disease has also been devastating, causing an economic loss of nearly $12 billion annually. The significant impact of this loss has been well felt in developing countries, as it expands their poverty rate and margin among its populace. As a means to preserve the public health, early detection and analysis of bacterial pathogens which could be life-threatening is quite essential [10]. These could be a landmark achievements in clinical medicine, agriculture, food safety, public health and biosecurity. Pathogen detection methods is classified into the following sections: conventional methods, mass spectroscopy, and sensor-based methods.

2. Conventional Methods for Pathogen Detection

The detection of pathogens using conventional methods depends mostly on strategies which involve precise identification based on microbiological, biochemical
(phenotypic) and molecular (genotypic) constituents displayed by the organisms [11]. Sub-methods described in most conventional methods include traditional-based methods, immunology-based methods and nucleic acid-based methods [12,13,14]. Traditional-based method otherwise termed culture-based avails several approaches for pathogen detection. This method depends on the culturing of microbes on agar plates, the most probable number, membrane filtration, and many more. They are a widely recognized approach for their low cost and ease of use. The culturing techniques are highly dependable, relatively interesting compared to other methods, and expressing results both qualitatively and quantitatively [15]. Qualitative traditional-based methods determine the presence or absence of pathogens in samples, while quantitative traditional-based methods are quite useful for enumeration. Immunology-based method has been the most popular, successful and widely accepted technology in bacterial, spores, viruses and toxins detection, especially for Gram-negative bacteria. Several techniques associated with immunological detection include enzyme immunoassay (EIA), enzyme-linked fluorescent assay (ELFA) [16,17], enzyme-linked immunosorbent assay (ELISA) [18], flow injection immunoassay, and others. It is faster (require less time preparing assay than traditional-based methods), robust and possesses the ability to detect organisms responsible for the contamination. ELISA and lateral flow immunoassay are the most accepted immunological methods applicable in foodborne pathogen detection, as they are not laborious and a large number of samples can be analyzed [19,20].

In the use of molecular based methods for the detection of pathogens, several techniques embedded in the nucleic acid-based method includes polymerase chain reaction (PCR), 16S rRNA (Denaturing Gradient Gel Electrophoresis-DGGE, Restriction Fragment Length Polymorphism-RFLP, and Ribosomal RNA Intergenic Spacer Analysis-RISA), Fluorescence in-situ Hybridization-FISH, microarray, Sequencing, Next Generation Sequencing (NGS) and many more. These methods have proven its efficiency by preventing ambiguity as well as wrong interpretation of results. Due to their precision and accuracy, they become imperative and are categorized as conventional techniques for the detection of pathogens. The nucleic acid-based method showed high sensitivity and specificity as it involves the use of high-throughput techniques to identify microbes, address their sources and point of variations, and thus, revolutionizing approaches to the study of microbial and clonal diversity.

2.1. Mass Spectroscopy for Pathogen Detection

A new phase towards the detection of pathogens was achieved with the advent of Matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry (MS). These techniques identify microorganisms based on the uniqueness of the mass protein spectral fingerprints present on their cell surfaces. It has been applied for the detection of pathogens protein marker signals in different samples [21]. One significant advantage of MALDI-TOF MS is the dynamism of its proteome profiling property which enables the addition of new microorganism’s protein spectral into the database [22]. However, few setbacks in this method include the need for enrichment step before a test, the availability of the peptide mass fingerprints in the database which must correspond and identify tested microorganisms, high cost of procurement and setting-up.

2.2. Limitations of the Established Pathogen Detection Methods

In spite of the benefits derived from each method, some major setbacks limiting its adequacy for point and quick detection approach has not been pleasing well enough. Consideration of the conventional methods, shows they are laborious -taking 2-3 days and 6-7 days often before initial results and specific pathogens are determined respectively. In some cases, the futility of normal culture plate technique often requires pre-enrichment, selective enrichment, selective plating and identifications. The sequence of the method is stereotyped, lengthy, time-consuming, limited by its low sensitivity -giving false-negative results even with the presence of viable but non-culturable (VBNC) cells [23,24]. Immunology-based methods present varying disadvantages such as the use of expensive test kits, false result due to cross-reactivity with closely related antigens, the need for pre-enrichment to attain detectable antigen level in samples. Major difficulties are also often encountered when expert staff default in engaging proper laboratory procedures which include proper labelling of antigens and antibodies, and accommodating external interference that should be limited. Its dependence on the amount of antigen in the sample which determines the specific binding response from the antibodies is also of major concern [25].

Furthermore, PCR methods - a highly predictive method still requires the use of expensive instruments for its nucleic acid amplification and quantification. Other complexities of PCR are the necessities for specific primers after the microorganisms’ subjecting to probable cultural identification, its optimal reaction mixture which requires trained personnel to operate and avoid risk of false positive and negative results. Sensor-based method devised for pathogenic substances detection without special sample pre-enrichment, is still limited due to the difficulty experienced in the enhancement of immobilized bio-components stability. Its high cost in instrumentation design and quality assurance slows its commercial and laboratory methods than other rapid methods [25]. These setbacks highlighted above creates the need to search and re-evaluate existing methods for newer approaches affordable and efficient for pathogen detection.

3. Sensor-Based Methods for Pathogen Detection

Sensor-based methods have been proven to be a dependable device useful for the detection of living organisms, biological molecules as well as to detect chemical components present in living organisms. Recently, research activities have increasingly drifted towards the use of the sensor-based approach for pathogens detection. They have become essential in pathogens and toxins
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Detection, environmental monitoring [26], soil quality monitoring, drug discovery, prosthetic devices [25], timely detection of post-harvest deterioration [27], and food quality monitoring [28]. The wide applications are as a result of its short-time analysis, portability in the design of biological analytical techniques, exemption of sample pre-enrichment approach, its efficiency in real-time measurements and automation [11]. The use of a sensor-based method has become a favorable method to ensure food safety both in real-time and during the production process [29]. Food industries are currently engaged in the development/use of bio-recognition elements for pathogen detection in products. Sensor-based methods which had gained rapid technological growth is constituted of two major materials namely the bio-receptor (biological capture molecule/biosensing or bio-recognition elements) and the transducer -which converts bio-recognized energy/biological response into signals.

Bio-receptors are an important component in the development of sensor-based methods because their surfaces are specifically designed using some biochemical mechanisms to recognize and initiate binding with the analyzed materials. Varying bio-receptors reported for sensor-based method efficiency include antigens, antibodies, enzymes, nucleic acids, cells, microorganisms, aptamers, bio-mimic substance and many more as shown in Figure 1. Binding initiation by the bio-receptors to the analyzed materials of interest activates the reaction for sensor measurement via a transducer [30].

Optical, electrochemical, spectroscopy and magnetic capture/separation transducers were found to be widely used for pathogen detection. Similarly, some of the widely accepted transducers have also been reported to be used in combination with other less important ones for improved efficiency [31,32,33].

3.1. Functionalizing Nanoparticles for Pathogen Detection in Sensor-based Methods

Improving the sensitivity of bio-receptors for bacteria detection ushered acceptance in the use of nano-materials as signal amplifiers. Functionalizing nano-materials with bio-receptors showed a high level of success in the development of bacterial biosensors due to its good conductivity, high surface-to-volume ratio, diffusion rate and good result outputs on the transducers [5]. The feat is achievable because nanotechnology through its approach encourages bio-fabrication of useful devices from nano-materials to form conjugate macromolecules with other materials. Yang et al. [36] in their study on the subject established the advantages of incorporating nanoparticles in biosensor which includes rapid and real-time detection, ensure an improved detection sensitivity and enabling the detection of multiple pathogens simultaneously. Also, they are versatile, easy to manipulate, biocompatible enough to enhance signal effects when combined with varieties of biological, molecular and artificial/synthetic materials, and thus, produces a visible colorimetric result [5].

Figure 1. Essential components in Biosensor-based methods (Adapted from 19,34,35)
Surface modification and functionalizing inorganic nanoparticles (metal and semi-conductor) with bio-recognition elements have become a widespread practice with many attempts to improve and ensure its analytical sensitivity [37]. Metal and semi-conductor nanoparticles (Table 1) functionalized with materials provides amplified signals for the detection of bacteria [38]. For proper functionalization and modification, appropriate bio-receptors materials are required to provide active biocompatibility with analyzed materials, enhanced signals as well as rapid detection procedure [39]. Thus, in achieving selectivity and improve sensitivity, nano-materials needs to be interfaced with biological, molecular and artificial receptors for specific binding and target of bacteria [40].

### Table 1. Forms of Inorganic Metallic Nanoparticles

<table>
<thead>
<tr>
<th>Forms</th>
<th>Metals Involved</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pure Metals</td>
<td>Ag, Au, Cu, Fe, Ni, Co, Pt, Palladium</td>
<td>[41,42]</td>
</tr>
<tr>
<td>Metal oxides (Semi-conductor)</td>
<td>ZnO, CuO, TiO₂, CrO₂, SiO₂</td>
<td>[41,42]</td>
</tr>
<tr>
<td>Magnetic</td>
<td>FeO₄, Fe₂O₄</td>
<td>[42]</td>
</tr>
<tr>
<td>Metallic</td>
<td>PbS, ZnS, CdS, FeS, HgS, ZnSe</td>
<td>[42]</td>
</tr>
<tr>
<td>Chalcogenides</td>
<td>CdTe, CuInSe, Ag-Au; Zn-Ag, Pt-Ni, Co-Mo, Ag-MgO, FeO₂-Au, Fe₂O₄-Ag, MnFe₂O₄-Au, Fe-Pt, Fe₃O₄-Zn, Fe₂O₃-Zn</td>
<td>[2,42-46]</td>
</tr>
</tbody>
</table>

Advancement in nanotechnology and its approach towards the exploration of more sensitive optical biosensors had created more interest in the development of new nano-structured materials, especially metal for improved overall performance as bacterial biosensors [26,47,48]. Attachment of bio-receptors to the surfaces of nano-materials (Figure 2) have been either through a direct or indirect method. Direct method namely: physical adsorption or covalent coupling could both be exhibited by hydrophobic and electrostatic interactions. Polyethylene glycol (PEG), Poly-L-lysine (PLL), and Polyethylenimine (PEI) are typical examples of materials used. The indirect method forms a bridge link between biomolecules and nano-materials with corresponding high affinity with biotin, avidin, and streptavidin as typical examples of linkers [49]. According to Yang et al. [36], the conjugation of biomolecules with nano-materials is the foundation of nano-biorecognition. Based on the nature of biomolecules conjugated to nano-materials, there are antibody-antigen, adhesin-receptor, antibiotic-antigenic surface, and complementary DNA sequence recognitions.

#### 3.1.1. Functionalized Pure Noble Metal Nanoparticles (Gold Nanoparticles)

A novel attempt which involved the use of monomeric sugar to label a protein present on the bacteria cell surface to aid microscopy view has been achieved. Lin et al. [50] reported functionalized gold nanoparticles (AuNPs) with monomeric mannose sugar specifically recognized adhesion FimH of type 1 pili in Escherichia coli. It has been documented that antibody tends to produce reliable affinity, increases surface area to volume ratio and also amplifies signals. The use of a specific antibody as functionalizing agent for AuNPs was reported by Basu et al. [51], with preferences given to the development of immunochromatographic strip (Figure 3) for the detection of Salmonella typhi [52,53]. Other work by Baccar et al. [54] reported the use of acid-thiol, amine-thiol coupled with antibody to surface-modified gold nano-materias. Pengsuk et al. [55] in their study established the detection of Vibrio cholerae 0139 in seafood using gold nanoparticles and specific antibodies.

![Figure 2. Overview of bacterial detection by functionalized inorganic nanoparticles](image1)

![Figure 3. Components of an immunochromatographic strip](image2)
Augmenting antibody with other bio-receptors such as chitosan to serve as cross-linker effectively increases surface area for reactivity as a result of their mesh structure. The combined effect of antibody-chitosan carried out by Kang et al. [56] showed the limit of detection as 10 CFU/mL. Thiramanas and Lacharoensuk [57] showed the effectiveness of Polyethyleneimine-AuNPs to electrostatically bind to the surfaces of enterotoxigenic E. coli (ETEC) and S. aureus. PEI increases and stabilizes the electrostatic interaction (direct method) between the positively charged gold nanoparticles and negatively charged components on bacteria cell surfaces. Findings by Raj et al. [58] and Huang et al. [59] showed the use of cysteine and 4-mercaptophenylboronic acid (4-MPBA) as a good functionalizing agent respectively. This direct method of modifying AuNPs showcase color responses which can be directly observed with the naked eye or through digital-camera-based red, green and blue (RGB) colour model analysis. Aggregation of DNA probes on the surface of AuNPs was reported for the detection of Mycobacterium sp. and S. aureus [60].

On the other hand, surface modification of AuNPs with aptamers gained lots of attention in the detection of Salmonella typhi and Shigella flexneri in research works [61,62,63,64,65]. The major advantage of Aptamers is in its ability to prevent AuNPs from NaCl-induced aggregation. It was found to increase electron transfer, electrochemical signal and ascertain detection within a short period (20-60 minutes). Few studies evaluated the bi-functional approaches of aptamer with other bio-receptors such as 4-mercaptobenzoic acid (4-MBA) [66], oligonucleotide probe [67], chitosan [68], 4-MBA and citrate [69]. Each complementing bio-receptors reported showed fast, sensitive, specificity and accurate SERS based sensors towards the detection of pathogens in samples. The incorporation of 4-MBA tags as shown in these studies specifically increases Raman signal. Varying forms of these bioreceptors are shown in Table 2.

3.1.2. Functionalized Magnetic Nanoparticles

Prior to the engagement of magnetic nanoparticles in pathogen detection, a simple approach to separating and concentrating charged targets in aqueous solutions has been provided [5]. Magnetic nanoparticles (MNPs) - a derivative of complex iron salts, have gained thorough studies and widespread applications especially for pathogen detection due to their stability, biocompatible and large surface-to-volume ratio aiding microbial cell wall adherence for effective separation [70,71]. Most importantly, they tend to become more useful and beneficial with their readiness to combine and improve detection platforms such as mass spectra, surface plasmon resonance, electrochemical, Raman spectra, fluorescent, and many more [72] as shown in Table 3. Ju [73] in his review expatiated the use of Fe₃O₄ nanoparticles-based hybridized materials to enhance peroxidase-like activity, and thus enabling the detection of bio-materials.

Lin et al. [74] evaluated the importance of vancomycin as an active functionalizing component for MNPs showing some level of compatibility with Gram-positive bacteria as it analyzes S. aureus and S. saprophyticus. Vancomycin inclusion limits MALDI-TOF MS false error as it reduces the interference of protein and metabolite signals in the mass spectra of Gram-positive bacteria. Vancomycin is a target antibiotic with high specificity for D-Ala-D-Ala moieties on Gram-positive cell walls. Several studies documented the use of varying binding materials to attach antibodies to the surfaces of MNPs to establish high adherence, biocompatibility and rapid trapping of targeted pathogens [75,76,77]. The inclusion of HRP to antibody functionalized MNPs by Mun and Choi [78], prevents false positive signals. The development of DNA aptamer-coated MNPs for pathogen detection showed a realistic approach by improving peroxidase-like activities of MNPs and thus increases signals. Surface enhancement of MNPs with aptamer as described by Park et al. [79] and Wang et al. [80] showed improvement in the detection sensitivity of S. typhi and S. aureus. The application of oligonucleotide probe as functionalizing bio-element of MNPs was addressed by Li et al. [81] to aid instant detection of their bacterial targets.

Different approach explored by Matta and Alocilja [82] maximizes carbohydrate ligands (gycan and cysteine-gycan) as the functionalizing component for the detection of S. enteritidis, E. coli O157:H7 and B. cereus in pasteurized milk. As earlier documented, non-covalent electrostatic interaction towards bacteria cell surface is due to the presence of hydroxyl, amino and hydrophobic regions present in carbohydrate ligands. Functional groups on ligands show high affinity for lipids, sugars and proteins present on bacteria cell surfaces [83,84]. Functionalized MNPs by urease enzyme hydrolysis urea, increases the pH value and enhances the binding strength of the complex [72]. Hydrolysis of urea by urease enzyme in this work report colorimetric detection of the pathogen as a result of the increase in pH and thus promote detectability on litmus dye. Recent research articles showed the potential of chitosan. Le et al. [85] suggested that protonated amine group on chitosan attracts efficient and stronger binding quality with the anionic components on bacteria surfaces (E. coli and S. aureus) in an acidic pH condition.

3.1.3. Functionalized Metal Oxides Nanoparticles

Studies have documented the potential of metal oxides surface modification for capturing and detection of pathogens [87,90]. Amongst metal oxides, silica oxides (SiO₂) enables high photo-stability and conjugation with biomolecules. Zhao et al. [86] and Wang et al. [87] in their study reported functionalizing silica nanoparticles (SiO₂NPs) surfaces with antibodies for the detection of E. coli O157:H7, S. typhi, and S. aureus (Table 4). It was shown that the procedure facilitates strong and specific antibody-antigen interaction and recognition. Study on titanium oxide (TiO₂) revealed it has a wide band gap semi-conductor and good stability [88,89] which shows its essentiality for sensor-based detection. Viter et al. [90] evaluated the importance of titanium oxide nanoparticles (TiO₂NPs) functionalized by antibody and deposited on glass substrates for the detection of Salmonella. In their research findings, antibody-TiO₂NPs conjugates exhibited intense photoluminescence at the visible range spectrum which was attributed to the strong electrostatic interaction between TiO₂NPs and the antibody protein.
Table 2. Some Bacteria Detected in Functionalized Gold Nanoparticles (AuNPs)

<table>
<thead>
<tr>
<th>Target Organisms</th>
<th>Bioreceptor Used</th>
<th>Developed Mechanisms</th>
<th>Detection Methods</th>
<th>Limit of Detection (CFU/ml)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>E. coli</td>
<td>Mannose</td>
<td>Coupling of monomeric mannose to gold nanoparticle (m-AuNps) to target adhesin FinH of type 1 pili of the target pathogen. Immobilized Gold nanowire arrays (GNWA) bounded with antibody prepared on alumina template trapped target pathogen to form a sandwich with second antibody conjugated with phosphatase.</td>
<td>Microscopy (TEM)</td>
<td>NA</td>
<td>[50]</td>
</tr>
<tr>
<td>E. coli 0157:H7</td>
<td>Antibody</td>
<td>Acid-thiol modified gold substrate form a conjugated with polyclonal antibody gold nanoparticle. DNA probe of enzyme ALP and detector probe coated on gold nanoparticles were immobilized on indium tin oxide (ITO) electrode-coated glass slides.</td>
<td>Electrochemical Impedance Spectroscopy (EIS)</td>
<td>10 cells/0.173 cm²</td>
<td>[51]</td>
</tr>
<tr>
<td>E. coli K12; L. fermentum</td>
<td>Antibody</td>
<td>Antibody coated gold nanoparticles were clotted on nitrocellulose membrane to form an immunochromatographic strip.</td>
<td>Surface Plasmon Resonance (SPR)</td>
<td>10⁴ &amp; 10³</td>
<td>[54]</td>
</tr>
<tr>
<td>Mycobacterium sp.</td>
<td>DNA Probes</td>
<td>Antibody coated gold nanoparticles were clotted on nitrocellulose membrane to form an immunochromatographic strip.</td>
<td>Electrochemical Impedance Spectroscopy (EIS)</td>
<td>1.25 ng/ml</td>
<td>[60]</td>
</tr>
<tr>
<td>S. typhi</td>
<td>Antibody</td>
<td>Antibody coated gold nanoparticles were clotted on nitrocellulose membrane to form an immunochromatographic strip.</td>
<td>Colorimetric</td>
<td>1.14 x 10⁵</td>
<td>[52]</td>
</tr>
<tr>
<td>Salmonella; E. coli 0157:H7</td>
<td>Aptamer</td>
<td>Aggregation of aptamers on the surface of gold nanoparticles and signal amplification upon high salt conditions.</td>
<td>Colorimetric; Optical UV-Vis</td>
<td>10⁵</td>
<td>[61]</td>
</tr>
<tr>
<td>B. cereus</td>
<td>Antibody-Chitosan</td>
<td>Monoclonal antibody-modified gold nanoparticles trapped target pathogen was cross-linked to chitosan immobilized on glassy carbon electrode. Biotinylated aptamer immobilized on biotin and streptavidin-coated microtiter plate-wells trapped target pathogen, and sandwiched with aptamer-coated gold nanoparticles to complete the assay.</td>
<td>Amperometric (Cyclic voltammetry)</td>
<td>10</td>
<td>[56]</td>
</tr>
<tr>
<td>S. typhi</td>
<td>Aptamers</td>
<td>Gold nanoparticles were immobilized on nitrocellulose membrane to form a sandwich with second antibody conjugated with phosphatase.</td>
<td>Colorimetric; UV-Vis Spectroscopy @ 630 nm</td>
<td>7</td>
<td>[62]</td>
</tr>
<tr>
<td>E. coli 0157:H7</td>
<td>Cysteine</td>
<td>Gold nanoparticles were immobilized on nitrocellulose membrane to form a sandwich with second antibody conjugated with phosphatase.</td>
<td>Colorimetric</td>
<td>100</td>
<td>[58]</td>
</tr>
<tr>
<td>ETEC; S. aureus</td>
<td>Polyethylenimine</td>
<td>Monoclonal antibody-modified gold nanoparticles trapped target pathogen was cross-linked to chitosan immobilized on glassy carbon electrode. Biotinylated aptamer immobilized on biotin and streptavidin-coated microtiter plate-wells trapped target pathogen, and sandwiched with aptamer-coated gold nanoparticles to complete the assay.</td>
<td>Colorimetric; UV-Vis Spectroscopy @ 630 nm</td>
<td>7</td>
<td>[62]</td>
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<td>Colorimetric</td>
<td>100</td>
<td>[58]</td>
</tr>
<tr>
<td>Strep. agalatiae</td>
<td>Oligonucleotide probe; Aaptamer</td>
<td>Conjugate of monoclonal antibody and Gold nanoparticles were assembled on the nitrocellulose membrane to form an immunochromatographic strip and over laid with capturing antibody.</td>
<td>Colorimetric</td>
<td>1.5 x 10⁵</td>
<td>[53]</td>
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<tr>
<td>S. xyphi</td>
<td>Aaptamer</td>
<td>Monoclonal antibody-modified gold nanoparticles trapped target pathogen was cross-linked to chitosan immobilized on glassy carbon electrode. Biotinylated aptamer anchored on microtiter plate capture target pathogen and overlaid with 4-mercaptopentanoic acid-coated gold nanoparticles.</td>
<td>Colorimetric</td>
<td>4</td>
<td>[66]</td>
</tr>
<tr>
<td>S. typhi</td>
<td>Oligonucleotide probe; Aaptamer</td>
<td>Formation of Gold nanoparticles conjugated with bi-functional oligonucleotide probes and aptamer, and signal amplified upon NaCl solution addition</td>
<td>Colorimetric; UV-Vis Spectroscopy</td>
<td>10</td>
<td>[67]</td>
</tr>
<tr>
<td>S. flexneri</td>
<td>Aaptamer</td>
<td>Gold nanoparticles were immobilized on nitrocellulose membrane to form a sandwich with second antibody conjugated with phosphatase.</td>
<td>Colorimetric; UV-Vis Spectroscopy @ 400-800nm; LAMP</td>
<td>80</td>
<td>[65]</td>
</tr>
<tr>
<td>S. typhi</td>
<td>Carboxymethyl chitosan; Aaptamer</td>
<td>Composite comprising of carboxymethyl chitosan loaded with amino-modified aptamer functionalized gold nanoparticles to capture target pathogen.</td>
<td>Colorimetric; UV-Vis Spectroscopy @ 550nm</td>
<td>16</td>
<td>[68]</td>
</tr>
<tr>
<td>E. coli; S. pullorum; S. aureus; E. faecalis; S. mutans</td>
<td>4-mercaptophenylboronic acid</td>
<td>Gold nanoparticles modified with 4-MPBA aggregates on targeted pathogen surfaces.</td>
<td>Image capture (digital camera in black box); UV-Vis Spectroscopy @ 524nm</td>
<td>1.02 x 10⁸</td>
<td>[59]</td>
</tr>
<tr>
<td>Shigella sonnei</td>
<td>Citrate; Aaptamer</td>
<td>Citrate-stabilized Gold nanoparticles were conjugated with aptamer to target bacteria cells. Signal amplified with raman active 4-MBA ligand.</td>
<td>Surface Enhanced Raman Scattering (SERS) measurement</td>
<td>10</td>
<td>[69]</td>
</tr>
</tbody>
</table>
bio-functionality properties have been established [92,93]. Magnetic response, improved stability and multiple nanoparticles (larger than 100nm) producing a good Large nanocomposites of noble-metal and magnetic nanoparticles in the formation of nanocomposites. Bi-functional assessment of magnetic nanoparticles for the detection of pathogens. Two noble metals could co-exist as bi-metallic nano-materials for the detection of pathogens indicating that two noble metals could co-exist as bi-metallic nano-materials for the detection of pathogens. Interestingly, this was also the case of magnetic and metallic nanoparticles in the formation of nanocomposites [2]. Both magnetic and noble-metal nanoparticles have been studied and extensively applied as nanocomposites. Large nanocomposites of noble-metal and magnetic nanoparticles (larger than 100nm) producing a good magnetic response, improved stability and multiple bio-functionality properties have been established [92,93].

### Table 3. Some Bacteria Detected in Functionalized Magnetic Nanoparticles (MNPs)

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<thead>
<tr>
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<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>S. aureus; S. saprophyticus</td>
<td>Vancomycin</td>
<td>Magnetic nanoparticles were modified with vancomycin to selectively trapped target pathogens.</td>
<td>Immuno-magnetic separation (IMS); MALDI-TOF MS</td>
<td>7 x 10⁴</td>
<td>[74]</td>
</tr>
<tr>
<td>E. coli</td>
<td>Antibody</td>
<td>Biotinylated polyclonal antibodies were conjugated with streptavidin-coated magnetic nanoparticles to form a complex.</td>
<td>Immuno-magnetic separation (IMS); Plating</td>
<td>8</td>
<td>[75]</td>
</tr>
<tr>
<td>Listeria monocytogenes</td>
<td>Antibody</td>
<td>Magnetic nanoparticles modified with carboxylic acid and antibody.</td>
<td>Immuno-magnetic separation (IMS); Real-time PCR</td>
<td>226/0.5 ml</td>
<td>[76]</td>
</tr>
<tr>
<td>E. coli 0157:H7; S. enterica; V. cholera; C. jejuni; V. harveyi; S. aureus; B. cereus</td>
<td>Oligonucleotide probe</td>
<td>Streptavidin-coated magnetic nanoparticles hybridized with biotinylated oligonucleotide probe target bacteria cell.</td>
<td>Microscopy (CCD Camera)</td>
<td>316</td>
<td>[81]</td>
</tr>
<tr>
<td>S. typhi</td>
<td>Antibody</td>
<td>Magnetic nanoparticles conjugated with antibody was signal amplified upon the addition of horseradish peroxidase.</td>
<td>Immuno-magnetic capture</td>
<td>2 x10⁴</td>
<td>[77]</td>
</tr>
<tr>
<td>S. typhi</td>
<td>Monoclonal Antibody (MAb)</td>
<td>Monoclonal antibody coupled with magnetic nanoparticles form complexes with bacteria cell, and enhances trapping on nitrocellulose filter.</td>
<td>Colorimetric NA</td>
<td>NA</td>
<td>[79]</td>
</tr>
<tr>
<td>S. typhi</td>
<td>Aptamer</td>
<td>Development of magnetic nanoparticles modified with aptamers and optical signal amplified with 3',3',5,5'-tetramethylbenzidine (TMB) in the presence H₂O₂.</td>
<td>Colorimetric</td>
<td>10⁴ (Naked eye</td>
<td>[85]</td>
</tr>
<tr>
<td>S. enteritidis; E. coli 0157:H7; B. cereus</td>
<td>Carbohydrate ligands</td>
<td>Glycan and cysteine-glycane were aggregated on magnetic nanoparticle for microbe extraction.</td>
<td>Magnetic capture</td>
<td>2.19 in B. cereus</td>
<td>[82]</td>
</tr>
<tr>
<td>S. aureus; A. jundi; V. harveyi; M. luteus</td>
<td>Urease</td>
<td>Quanternized magnetic nanoparticles optical signal amplified with urease, urea solution and phenol red.</td>
<td>Colorimetric; Microplate reader @ 558nm</td>
<td>10²</td>
<td>[72]</td>
</tr>
<tr>
<td>E. coli; S. aureus</td>
<td>Chitosan</td>
<td>Positively charged chitosan were coated on magnetic nanoparticles and signal amplified with 2,2’-azino-bis(3'-ethylbenzothiazoline-6-sulphonic acid) (ABTS) in the presence of H₂O₂.</td>
<td>Colorimetric</td>
<td>10² (Spectroscopy)</td>
<td>[85]</td>
</tr>
</tbody>
</table>

### Table 4. Some Bacteria Detected in Functionalized Metal oxides Nanoparticles

<table>
<thead>
<tr>
<th>NPs used</th>
<th>Target Organisms</th>
<th>Bioreceptor Used</th>
<th>Developed Mechanisms</th>
<th>Detection Methods</th>
<th>Limit of Detection (CFU/ml)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>SiO₂NPs</td>
<td>E. coli 0157:H7</td>
<td>Antibody</td>
<td>Attachment of monoclonal antibodies on surfaces of RuBpy-doped silica nanoparticles.</td>
<td>Microscopy (Fluorescent); Flow cytometry</td>
<td>1 cfu/g</td>
<td>[86]</td>
</tr>
<tr>
<td>E. coli; S. typhi; S. aureus</td>
<td>Antibody</td>
<td>Co-encapsulation of tandem dyes for signal amplification on silica nanoparticles conjugated with monoclonal antibodies.</td>
<td>Microscopy (Confocal)</td>
<td>NA</td>
<td>[87]</td>
<td></td>
</tr>
<tr>
<td>TiO₂NPs</td>
<td>Salmonella</td>
<td>Antibody</td>
<td>Deposition of monoclonal antibodies coated titanium oxide on glass surface to trap target pathogen.</td>
<td>Photoluminescence</td>
<td>10⁵</td>
<td>[90]</td>
</tr>
</tbody>
</table>

3.1.4. Functionalized Bi-Metallic Nanoparticles

The formation of unique nanocomposites with the combination of two metals suggests their potential and importance as composite for the detection of pathogens (Table 5). Duan et al. [91] conducted a sandwich-like detection strategy functionalizing Au-Ag nanocomposites with aptamer and X-rhodamine (ROX)-modified aptamer indicating that two noble metals could co-exist as bi-metallic nano-materials for the detection of pathogens.

Interestingly, this was also the case of magnetic and metallic nanoparticles in the formation of nanocomposites [2]. Both magnetic and noble-metal nanoparticles have been studied and extensively applied as nanocomposites. Large nanocomposites of noble-metal and magnetic nanoparticles (larger than 100nm) producing a good magnetic response, improved stability and multiple bio-functionality properties have been established [92,93].
magnetic nanocomposites (SiO2-MNPs) has shown higher capturing efficiency. Ji et al. [99] discussed the use of silica-coated magnetic nanoparticles, with manganese (Fe3O4). Amagliani et al. [100] established the use of oligonucleotide probe to surface modify SiO2-MNPs. Other work assessed D-mannose-lectin and 3-Aminopropyltriethoxysilane as the functionalizing bio-elements respectively [70,101]. The method showed a relatively good magnetic response and microscopy detection with the use of fluorescein-labeled Concanavalin A, identifying the possibility of functionalized/enhanced silicon shell for magnetic nanoparticles. Similarly, Bai et al. [102] in their study validated the use of non-noble metal (silica) as the coating shell for MNPs (Fe3O4). Amagliani et al. [100] established the use of oligonucleotide probe to surface modify SiO2-MNPs. Other work assessed D-mannose-lectin and 3-Aminopropyltriethoxysilane as the functionalizing bio-elements respectively [70,101]. The method showed a relatively good magnetic response and microscopy detection with the use of fluorescein-labeled Concanavalin A, identifying the possibility of functionalized/enhanced silicon shell for magnetic nanoparticles. Similarly, Bai et al. [102] in their quest for the detection of Acinetobacter baumannii focused their novel mechanism on the use of bacteriophages tail-fibre protein -TF2 and TF6, as the functionalizing bio-element for alumina-coated magnetic nanoparticles. This authenticates the peculiarity of tail fibres attachment of bacteriophages to their bacteria host cells.

Table 5. Some Bacteria Detected in Functionalized Bi-metallic Nanoparticles (Nanocomposites)

<table>
<thead>
<tr>
<th>NPs used</th>
<th>Target Organisms</th>
<th>Bioreceptor Used</th>
<th>Developed Mechanisms</th>
<th>Detection Methods</th>
<th>Limit of Detection (CFU/ml)</th>
<th>References</th>
</tr>
</thead>
<tbody>
<tr>
<td>E. coli</td>
<td>Polyclonal Antibody</td>
<td>Formation of biotinylated polyclonal antibodies on avidin-modified, gold-coated magnetic nanoparticles traps target pathogen and signal amplified with 5,5-dithiobis-(2-nitrobenzoic acid) coated gold nanorods.</td>
<td>Immuno-magnetic separation (IMS); Surface Enhanced Raman Scattering (SERS) measurement</td>
<td>8</td>
<td>[94]</td>
<td></td>
</tr>
<tr>
<td>E. coli BL21; S. aureus</td>
<td>Polyethyleneimine (PEI)</td>
<td>Formation of polyethyleneimine-modified gold-coated magnetic nanoparticles conjugates for effective magnetic capturing response</td>
<td>Magnetic capture; Surface Enhanced Raman Scattering (SERS) measurement</td>
<td>10^3</td>
<td>[2]</td>
<td></td>
</tr>
<tr>
<td>S. aureus</td>
<td>Polyethyleneimine (PEI)</td>
<td>Formation of polyethyleneimine-modified gold-coated magnetic nanoparticles fused with manganese</td>
<td>Immuno-magnetic separation (IMS); SERS measurement</td>
<td>10^9</td>
<td>[45]</td>
<td></td>
</tr>
<tr>
<td>MNPs &amp; AuNPs</td>
<td>Salmonella choleraesuis</td>
<td>Monoclonal Antibody</td>
<td>Monoclonal antibody-modified, gold-coated magnetic nanobeads were immobilized on Immunochromatographic strip pad.</td>
<td>Colorimetric</td>
<td>5</td>
<td>[95]</td>
</tr>
<tr>
<td>S. aureus</td>
<td>Aptamer; Vancomycin SERS tag</td>
<td>Dual recognition method involving the use of Aptamer-modified gold-coated magnetic nanoparticles and Vancomycin SERS tagged-coated AuNPs</td>
<td>Microscopy (TEM); SERS measurement</td>
<td>3</td>
<td>[97]</td>
<td></td>
</tr>
<tr>
<td>Group A Streptococcus pyogenes</td>
<td>Antibody</td>
<td>Collection of target pathogen using antibody-modified, gold-coated magnetic nanoparticles and signal amplified with Pyrrolidonyl arylamidase (PYR) and drops of 4-(dimethylamino)-(innamaldehyde (DMACA)</td>
<td>Colorimetric (Visual &amp; Image Analysis); Microscopy (SEM)</td>
<td>3.3 x 10^2</td>
<td>[96]</td>
<td></td>
</tr>
<tr>
<td>MNPs &amp; AgNPs</td>
<td>E. coli BL21; S. aureus 04018; MRSA</td>
<td>Vancomycin; Polyethyleneimine (PEI)</td>
<td>Development of the combination of polyethyleneimine and vancomycin-modified silver-coated magnetic nanoparticles and silver-gold nanoparticle conjugates.</td>
<td>Magnetic separation; SERS measurement</td>
<td>5 x 10^3</td>
<td>[98]</td>
</tr>
<tr>
<td>Listeria monocytogenes</td>
<td>Oligonucleotide probe</td>
<td>Fusion of oligonucleotide probe of selected pathogen gene with silica-coated magnetic nanoparticles.</td>
<td>Immuno-magnetic separation; PCR</td>
<td>10</td>
<td>[100]</td>
<td></td>
</tr>
<tr>
<td>MNPs &amp; SiO2NPs</td>
<td>E. coli</td>
<td>Mannose</td>
<td>Mannose binding lectin was immobilized on silica-coated magnetic nanoparticles and signal amplified with fluorescein-labelled concanavalin A.</td>
<td>Microscopy (Fluorescent)</td>
<td>10^4</td>
<td>[70]</td>
</tr>
<tr>
<td>S. enteritidis; Listeria monocytogenes</td>
<td>3'-Aminopropyltriethoxysilane</td>
<td>Magnetic extraction of target bacteria cell DNA using amino-modified silica-coated magnetic nanoparticles complex.</td>
<td>Magnetic extraction; PCR</td>
<td>8 (SE)</td>
<td>[101]</td>
<td></td>
</tr>
<tr>
<td>MNPs &amp; Al</td>
<td>Acinetobacter baumannii</td>
<td>Tail fibre protein of bacteriophages (TF2 and TF6)</td>
<td>Formation of TF2&amp; TF6-modified alumina-coated magnetic nanoparticles complexes.</td>
<td>MALDI MS</td>
<td>10^4 &amp; 10^5</td>
<td>[102]</td>
</tr>
<tr>
<td>MNPs &amp; PnNs</td>
<td>E. coli</td>
<td>Vancomycin</td>
<td>Attachment of vancomycin to the surface of platinum-coated magnetic nanoparticle (FePt).</td>
<td>Immuno-magnetic separation; Microscopy (SEM; TEM)</td>
<td>180</td>
<td>[103]</td>
</tr>
<tr>
<td>Au-Ag</td>
<td>S. typhi</td>
<td>Aptamers</td>
<td>Formation of sandwich assay comprising of primary aptamer-modified gold-silver nanocomposite, target pathogen and X-rhodamine-modified secondary aptamer.</td>
<td>SERS measurement</td>
<td>15</td>
<td>[91]</td>
</tr>
</tbody>
</table>
Platinum-coated magnetic nanoparticles (FePt) conducted on E. coli showed stability and solubility in water when modified with vancomycin (Van-FePt) [103]. Although, it is expected for vancomycin to show greater affinity for Gram-positive bacteria, yet, low concentration binding was observed in E. coli (Gram-negative bacteria).

On the other hand, surface modification and conjugation of different shape of inorganic metals had received some attention. Sandwich assay complex involving the aggregation of specific monoclonal and polyclonal antibodies on the surface of magnetic nanobeads and AuNPs respectively detect Listeria monocytogenes and S. typhi [104,105]. Wu et al. [106] examined the effect of aptamer-modified magnetic microparticles linked with antibody-coated gold nanoparticles. In contrast, a sandwich complex involving two aptamers (primary and secondary) was assayed by Abbaspour et al. [107] on magnetic beads and AgNPs. The findings showed the magnetic beads act as the carrier of the affinity ligands, ensuring fast magnetic separation. The two methods advocate their importance to colorimetric, immune-magnetic separation (IMS) and electrochemical detection of their respective target cells.

Huge difference to the previously described nanocomposites was the immobilization of aptamers as molecular recognition elements to surfaces of magnetic nanoparticles-coated upconversion nanoparticles (UCNPs). This was designed to efficiently capture and concentrate S. aureus, S. typhi and V. paraaemolyticus using the magnetic field and in the same vein amplified the luminescent signals [108,109].

Studies reviewed on nanocomposites established the importance and incorporation of iron oxide and magnetic nanoparticles as one of the bi-metallic components. This shows they possess fast and unique capturing properties but, also show a loss in reactivity level, specific surface area, porosity, dissolution rate, aggregation behavior, and stability after certain period. This deficiency is factored by its mineral structure [110,111]. To achieve effective applications of both iron oxide and magnetic nanoparticles, stable reactivity and longer shelf-life in different solutions is quite essential. The physical and chemical stability possessed by other pure and noble metals were observed to improve these defects in iron oxide and magnetic nanoparticles and does not affect their magnetization property when used as nanocomposites as shown in Table 6 [112].

### 4. Application of Functionalized Inorganic NPs for Pathogens Detection in Food and Water Samples

Food and water safety is an important measure that human and countries prioritize to certify their well-being [113]. The application of the bio-functionalized inorganic nanoparticles had in some ways improved the desire to safeguarding food products against contamination and all form of pathogen infestation. Few of the pathogens closely monitored both in developed and developing countries as the major food and water borne pathogens were practically detected via the application of functionalized inorganic nanoparticles assessed in this review. Seafood safety had received more attention due to the increase in their global demand [114]. Direct infestation, transmission and spread of these contaminated...
products by bacteria, biotoxins and other materials had consistently been reported in many countries [115,116]. Shrimps spiked with *S. aureus*, *V. parahemolyticus* and *S. typhimurium* were detected with AuNPs and MNPs-UCNPs functionalized with aptamers [67,109]. The co-existence of the three samples were reported to be efficiently and simultaneously detected in the samples. Wen-de et al. [53] achieved the detection of *Streptococcus agalatiae* injected in tilapia using an antibody and AuNPs-laced immunochromatographic strip. The finding reports a detection time of 15 minutes and further assured the validity of the strip for period of 4-6 months. AuNPs conjugated with aptamer application was verified on smoked salmon samples spiked with *Shigella flexneri* gaining a rapid detection time within 20 minutes of interaction [65].

In a similar attempt, attention was also drawn to pathogen detection in dairy products especially milk. Short shelf-life and easy contamination is mostly common in this product. Studies evaluating the detection of pathogens in spiked milk using biofunctionalized inorganic nanoparticles has been reported [68,76,91,95,100,101,106,117]. Investigation by Ma et al. [63] on the colorimetric detection of *S. typhi* using aptamer modified-AuNPs had 96.4% recoveries similar to those obtained using the plate counting methods (104.3%). Matta and Aloicijia [82] study which spanned for several months examined a total of 18 experiments as it considered the use of carbohydrate-functionalized MNPs. The study put into consideration environmental variations determining the proliferation of *S. enteritidis*, *E. coli* O157:H7 and *B. cereus* in milk samples. The capture efficiency of their analyses were reported to range between 73 to 90 %. Antibodies modified-nanocomposites detect the presence of *S. typhi* in milk spiked with two different concentration of the pathogen [105]. The developed method showed acceptable recovery values range of 91.5% and 106.8%.

Studies on artificial contamination of farm meat such as beef [75,86], pork meat [64,66] and chicken breast [69] were conducted as a case study to test its applicability for real time detection of pathogen in these products as well as similar products. Oh et al. [64] reported the usefulness of gold nanoparticle-aptamer-based LSPR sensing chip for *S. typhimurium* detection in pork meat. The study suggests an increase in the recovery rate as the proliferation of pathogen increases in the analyzed sample. The assessment of green products using MNPs have been reported. Chen et al. [104] illustrated the detection of *L. monocytogenes* in lettuce plants using antibodies functionalized nanocomposites (AuNPs and magnetic nanobeads). The feasibility of the method was established producing both colorimetric and optical reading at 588 nm. Similarly, Pang et al. [97] reported 50 minutes detection time for the detection of *S. aureus* in orange juice as it employs the use of inorganic nanoparticles bio-functionalized with an aptamer and antibodies. Antibiotics-SERS tag used improves the recovery rate from 95.0% to 106.4%.

The presence of ETEC, *E. coli* BL21 and *S. aureus* were determined with a polyethyleneimine-coated inorganic nanoparticles in artificially contaminated water [2,57]. The detection strategy showed rapid magnetic capture, visibility to the naked eye after 2-3 hours of interaction and optical readability within 10 minutes. Work by Huang et al. [59] reports gold nanoparticles modified with 4-MPBA produces both colorimetric and photographic detection outcome on 5 isolates (*E. coli*, *S. pullorum*, *S. aureus*, *E. faecalis*, *S. mutans*) within 20 minutes of analysis. Real water samples collected from lake, stream and puddle, without any form of pre-treatment were assessed for real-time detection of *E. coli*, *S. typhi*, *S. aureus* [94,108]. A comparison of the reported biofunctionalized methods with standard plate counting methods showed an average of 95.1% accuracy. Similarly, report by Abbaspour et al. [107] from real tap and river samples produced rapid results within 70 minutes of interaction with the aptamer modified inorganic nanocomposites.

### 5. Advantages of Inorganic Nanoparticles to Signal Amplification

Research findings widely recognized the usefulness of functionalized inorganic nanoparticles to pathogen detection. Several functionalizing strategies described has shown bright potential towards the improvement of biosensor methods for pathogen detection. Functionality gained had increased its use both in conventional and mass spectroscopy method of detection [118]. Conventionally, it improves assessment with the naked eye, absorbance quality and facilitates ionization of bacterial cell surfaces due to their adherence. However, the challenge of cost-effectiveness is still on the high side. Some nanomaterial-based approach still involved the use of expensive reagents and materials which are only available in the laboratory. Despite this challenge, move towards the development and use of simple materials such as paper-based (strip) methods and cotton buds [119] have gained much much. The approach and efficiency of strip methods to detect several analytes such as uric acid and glucose in body fluids influence the application towards the detection of a pathogen. This boosted the integration of paper-based method as a portable sensing and colorimetric detection device in food and water samples as seen in some of the reviewed studies. A typical example is the fabrication of an immunochromatographic strip, which has aided colorimetric detection mechanism and provides easier detection and assessment with the human naked eye. This technique has shown the possibility of its usefulness as a first-hand indicator in pool samples prior to the comprehensive subjection to complex laboratory screening for food and water samples. A patent technique (NG/PT/NC/2019/3865) established the importance of PLL-functionalized AgNPs as a colorimetric and optical detection tool on different strains of *E. coli* (EPEC, ETIEC, STEC and EAEC) within 2 hours of interaction. This investigation further adjudged functionalized inorganic nanoparticles-based detection as a promising, prospect-filled approach for rapid pathogen detection, and also informed that there may be little hurdles for its complete achievement, approval and acceptance.

Incorporation and combination of functionalized inorganic nanoparticles with molecular biological tools
such as DNA extraction and PCR reagents as reported in some reviewed studies is gradually enhancing its application in rapid, real-time and pathogen detection [120]. Other works by Li et al. [117] and Du et al. [121] documented that inorganic nanoparticles could still produce some colorimetric signals without binding to any bio-receptor. This suggests that direct attachment to PCR products with some concentration of salt solution ensures the accuracy of PCR products by eliminating false-positive results.

6. Conclusion

The aggregation of biomolecules on nanoparticles functionalizes and increases their binding affinity to target cells. Furthermore, it shows that the choice of an appropriate bio-recognition element is vital for rapid pathogen detection even at a low level. Therefore, the incorporation of suitable bio-receptors to functionalize these inorganic nanoparticles pave the way to the development of reliable miniature and more affordable tools able to detect food and waterborne pathogens. It is noteworthy that the quest for nanotechnology in the detection of the foodborne pathogens has drastically improved. Nevertheless, there is a still need to improve more on the known strategies which would readily detect both indicator and non-indicator but harmful microorganisms. The food and water industry should embrace, support and provide the needed support that will refine innovative ideas on functionalized nanotechnology inclusion for the detection of pathogens and toxins in food and water products.

Competing Interest

The authors have no competing interests.

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Nanoscience and Nanotechnology Research

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