Lipid Contents, Fatty Acid Profiles and Nutritional Quality of Nine Wild Caught Freshwater Fish Species of the Yangtze Basin, China

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Abstract Owing to the favorable effects of essential fatty acids on human health, a great degree of interest on fatty acid profiles and nutritional quality of fish species have been of interest in the recent years. The present study investigated the lipid content, fatty acid profiles and nutritional quality in nine freshwater fish species from the Ganjiang River in the Yangtze basin, China. Results showed that total lipid content of the dorsal muscle was 0.59–2.2% and that was inversely linked to moisture content (r2 = 0.79, p < 0.01). Wide ranges of monounsaturated fatty acids (MUFA) (21.83 to 50.53%) and polyunsaturated fatty acids (PUFA) (19.43 to 45.60%) were found. The ratios of n-3 to n-6 PUFA ranged from 0.25 to 1.16 (p < 0.001). N-3 PUFA was dominated by eicosapentaenoic and docosahexaenoic acids varying from 2.77 to 15.11% of total fatty acids. Indices of atherogenicity and thrombogenicity (IA and IT) based on fatty acid compositions ranged from 0.36 to 0.52 and 0.36 to 0.64 in all species, respectively (p < 0.001). These results indicated fatty acid profiles were different among the fish species and the potential of these species as dietary source of essential fatty acids from the nutritional standpoint.

Keywords: lipids, polyunsaturated fatty acids, fish, nutritional quality, Ganjiang River


1. Introduction

In the recent years the beneficial effects of highly unsaturated fatty acids in human nutrition has been increasingly recognized. For example such effects include its impacts on health, growth, vision among other characteristics [1,2,3]. In addition it is also thought that access to sea foods and hence the high contents of polyunsaturated fatty acids (PUFA) to our ancestors help develop the human brain, together with the acquisition of bipedalism made us what we are today [4,5]. PUFA, especially long-chain n-3 PUFA from fish or fish oil have demonstrated beneficial effects on clamping down several widespread diseases worldwide when consumed by humans according to epidemiological and statistical research [6,7].

Freshwater fish are generally characterized by low n-3 PUFA levels and high C: 18 PUFA levels compared to marine fish [8]. Fish, however, is one of the main sources of animal protein in the world, particularly among developing countries [9]. The high nutritional value of fish is also attributed to other nutritive materials and essential to humans such as minerals and vitamins [10]. Lipid contents and fatty acid compositions of fish species may differ even among different individuals of the same species. These variations result from geographical locations, dietary levels, feeding habits and other environmental factors [10,11,12,13]. However, freshness, availability, price and consumer preferences are important factors in the selection of fish for consumption [14,15]. Most individuals, especially residents of central and west China, consume freshwater fish as the availability and price of marine fish are relatively limited, whereas freshwater fishery and aquaculture in inland waters in China are well developed [9].

The fish species investigated in the present study are available in almost all markets nationwide and are important contributors to agricultural products exported. Although fish consumption is recommended in Chinese tradition due to its nutrients, the nutritional importance
2. Material and Methods

2.1. Sample Collection

A total of fifty-two samples of Carassius auratus, Cyprinus carpio, Pelleleobagrus fulvidraco, Megalobrama amblycephala, Siniperca chuatsi, Silurus meridionalis, Channa argus, Culter mongolicus and Channa asiatica representing five families were caught by local fishers from the Ganjiang River in the Yangtze basin, were randomly collected in May 2013. The number of each fish species analyzed and the mean-total length and somatic weight of all the specimens were recorded prior to the removal of the muscle samples for analysis. The freshly harvested fish were scaled immediately using a stainless steel knife. Skinless dorsal muscle samples were removed from the main edible portion of the fish and maintained in liquid nitrogen. All collected samples were moved after arriving at the laboratory and frozen at −80 °C prior to lipid extraction.

2.2. Lipid Extraction and Moisture Analysis

Moisture content was determined by drying the samples in an oven at 105°C to constant weight according to AOAC method [18]. Individual muscle samples measuring approximately 2.5 g were cut into small pieces before lipid extraction. The extraction process followed the method utilized by Bligh and Dyer [19] with minor modifications. The samples were saturated with 12.5 ml of chloroform/methanol/distilled water (2:2:1) containing 0.01% butylated hydroxytoluene. Organic liquid phase was obtained after mixed-phase separation. The liquid was passed through Whatman No. 1 filter paper and dried with moderate anhydrous sodium sulfate, then concentrated by using an evaporation-condensation rotating apparatus in a 40 °C water bath. Total lipid content was gravimetrically quantified. The moisture and lipid contents were expressed in g kg⁻¹ wet weight fish muscle.

2.3. Fatty Acid Analysis

Fatty acids were analyzed in the form of fatty acid methyl esters (FAMEs) prepared from the extracted lipid. The lipid extract of each individual fish was saponified with 0.5 M sodium hydroxide in redistilled absolute methanol at 70°C and then the fatty acids were methylated with 14% boron trifluoride methanol solution. Redistilled n-hexane was used in the extraction of derived FAMEs once the reaction mixture cooled to room temperature. The FAMEs were separated and quantified using GC-450 (Varian, USA) equipped with a flame ionization detector and an autosampler as described below. Individual FAME identification was performed by comparing the retention times of authentic mixture standards (Nu-Chek Prep, Inc., MN, USA) which were determined by GC-450/MS-320 (Varian, USA) with a DB-23 capillary column (0.25 mm i.d. × 0.25 μm film × 60 m length, J & W Scientific, USA). Oven temperature was programmed from 90 to 230°C. The injection port and detector temperatures were 270 and 250°C, respectively. Ultra-pure helium flowing at a rate of 1 ml min⁻¹ was used for each sample analysis and splitless injection of 1 μl sample was performed. The profiles of individual fatty acids were calculated using an automatic integrator and presented as percentages of total fatty acids according to the peak areas. Fatty acids in wet weight muscle were quantified according to the conversion factor of total fatty acids to total lipid content in finfish derived by [20].

2.4. Lipid Quality Indices

Indices of lipid quality in the fish species were estimated according to the method used by Ulbricht and Southgate [21] based on fatty acid compositions. Indices of atherogenicity (IA) and thrombogenicity (IT) were calculated by the following formulas:

\[
IA = \left[ \frac{(12:0) + (4 \times 14:0) + (16:0)}{(PUFA (n-6 and n-3) + MUFA)^{1/2}} \right]
\]

\[
IT = \left[ \frac{(14:0) + (16:0) + (18:0)}{(0.5 \times MUFA) + (0.5 \times n-6) + (3 \times n-3) + (n-3 \times n-6) + 1)} \right]
\]

2.5. Statistical Analyses

All of the data were presented as means ± standard deviation (SD). Statistical analyses of data were conducted using SPSS statistics software (version 17.0) for Windows. The data identified as non-homogeneous by Levene’s test were subjected to log transformation to confirm normal distribution and homogeneity of variance following comparison tests of experimental data prior to the statistical analyses. One-way analysis of variance (ANOVA) was used in the statistical comparisons. Fisher’s least significant difference (LSD) multiple-comparison tests were applied to the mean values of sample variances. Differences were considered significant when p < 0.05.

3. Results and Discussion

3.1. Total Lipid Content

Table 1 showed biometric data and biological indices of the fish collected. Total lipid content in the freshwater fish sampled varied greatly in this study (p < 0.05). C. argus...
had the lowest lipid content among the individual samples (4.3 g kg⁻¹) whereas *C. asiatica* had the highest lipid content (27.7 g kg⁻¹). All species, except *C. asiatica* were categorized as very low fat fishes (lipid content, < 2%) [22]. The mean lipid content of the nine fish species ranged from 5.9 to 22.0 g kg⁻¹ and averaged 11.3 g kg⁻¹. Two-thirds of the fish species were below 10.0 g kg⁻¹ in lipid content (Table 1). The low lipid levels were similar to those of wild freshwater fish from Lake Taihu in China which showed a smaller scope of the lipid contents, ranging from 5 to 13 g kg⁻¹ [17]. In contrast, Du et al. [16] reported that the lipid levels of freshwater fish from Chinese ponds had a wider range and a greater mean (9.5 to 38.5 g kg⁻¹ and 20.4 g kg⁻¹, respectively), meanwhile five freshwater fish species found in Indian ponds had slightly higher levels (6.0 to 25.5 g kg⁻¹ and 12.9 g kg⁻¹, respectively) than those in this study [23]. The results further confirmed the findings that wild fish generally have lower lipid levels than cultured fish [11,24]. However, some wild freshwater fish species from the Brazilian Pantanal exhibited high lipid contents [25]. In general, freshwater fish are typically characterized by low lipid content, although a plenty of researches have reported the effects of numerous factors on fish lipid content [26,27,28]. In addition, moisture content in the fresh muscles of the collected fish was notably high (Table 1). Our results revealed an inverse correlation between moisture content and total lipid ($r^2 < 0.79, p < 0.01$). Various studies on different types of fish reached a similar conclusion (Figure 1). Fish with relatively low moisture content had high lipid content. Moreover, this relationship was also linearly observed in farmed species and significantly in fatty fish [29].

### 3.2. Fatty Acid Profiles

Total fatty acid profiles in fish muscle were compared and listed in Table 2. Fatty acids were categorized into saturated fatty acids (SFA), which comprised 27.00 to 35.95% of total fatty acids in all fish species; monounsaturated fatty acids (MUFA), which accounted for 21.83 to 50.53%; and polyunsaturated (PUFA), which constituted 19.43 to 45.60%. Results revealed that the fatty acid composition profiles of the fish species investigated were markedly different. *C. auratus* had the highest PUFA content but the lowest MUFA. The opposite tendency was observed in *C. asiatica* and *C. mongolicus*. SFA variation was evidently smaller than those in MUFA and PUFA, although the three types of fatty acids in the present study displayed comparable mean levels (32.72, 34.47 and 32.81%, respectively). However, the range of MUFA in the current study was larger than those reported by previous studies [17,25]. Scatter diagrams indicating the relationships between total lipid content in fish muscle and SFA, MUFA and PUFA levels in total fatty acids were given in Figure 2. Noticeable changes in SFA percentage with lipid content variation were not observed. As shown in Figure 2a, the result was consistent with the previous result that SFA content was not greatly affected by lipid variations in fish muscle [29]. MUFA percentage tended to increase when total lipid content increased (Figure 2b), whereas a negative association between total lipid content and PUFA (Figure 2c). Similar conclusion was obtained by [31], although they found a different relationship between SFA percentage and total lipid content in Australian fish. Total lipids generally consist of various lipid classes, and the percentages of different fatty acid types are reflected in most lipid classes, and the proportions of different fatty acid types are reflected in various lipid compositions. Therefore, specific and individual lipid classes in fish should be investigated further to obtain additional knowledge regarding the correlation between total lipid content and fatty acid profiles.

### Table 1. Biometric data and biochemical indices of nine wild freshwater fish species

<table>
<thead>
<tr>
<th>Latin name</th>
<th>N'</th>
<th>Total length (cm)</th>
<th>Standard length (cm)</th>
<th>Somatic weight (g)</th>
<th>Moisture (g kg⁻¹)</th>
<th>Total lipid (g kg⁻¹)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>C. auratus</em></td>
<td>8</td>
<td>19.4 ± 4.3</td>
<td>15.9 ± 3.7</td>
<td>127.0 ± 57.4</td>
<td>803.0 ± 7.3</td>
<td>7.0 ± 3.0</td>
</tr>
<tr>
<td><em>C. carpio</em></td>
<td>7</td>
<td>33.2 ± 7.6</td>
<td>27.9 ± 6.5</td>
<td>765.3 ± 607.5</td>
<td>789.3 ± 20.7</td>
<td>7.8 ± 2.3</td>
</tr>
<tr>
<td><em>P. fulvidraco</em></td>
<td>5</td>
<td>25.4 ± 3.4</td>
<td>22.0 ± 3.0</td>
<td>156.7 ± 33.1</td>
<td>779.4 ± 14.0</td>
<td>16.7 ± 6.2</td>
</tr>
<tr>
<td><em>M. amblycephala</em></td>
<td>5</td>
<td>29.8 ± 4.2</td>
<td>25.7 ± 3.7</td>
<td>287.5 ± 162.6</td>
<td>784.8 ± 5.9</td>
<td>8.5 ± 1.4</td>
</tr>
<tr>
<td><em>S. chuatsi</em></td>
<td>5</td>
<td>23.4 ± 6.0</td>
<td>20.6 ± 5.1</td>
<td>180.2 ± 152.0</td>
<td>796.8 ± 14.5</td>
<td>7.1 ± 2.4</td>
</tr>
<tr>
<td><em>S. meridoualis</em></td>
<td>6</td>
<td>29.8 ± 4.9</td>
<td>27.0 ± 5.5</td>
<td>177.4 ± 58.1</td>
<td>793.8 ± 6.9</td>
<td>6.5 ± 2.0</td>
</tr>
<tr>
<td><em>C. argus</em></td>
<td>6</td>
<td>33.0 ± 4.7</td>
<td>28.3 ± 4.1</td>
<td>378.8 ± 160.4</td>
<td>791.5 ± 14.0</td>
<td>5.9 ± 1.2</td>
</tr>
<tr>
<td><em>C. mongolicus</em></td>
<td>4</td>
<td>43.9 ± 5.2</td>
<td>38.5 ± 4.9</td>
<td>1042.6 ± 334.6</td>
<td>778.5 ± 10.1</td>
<td>18.5 ± 3.4</td>
</tr>
<tr>
<td><em>C. asiatica</em></td>
<td>3</td>
<td>28.6 ± 1.3</td>
<td>24.6 ± 1.1</td>
<td>191.4 ± 39.6</td>
<td>763.7 ± 5.9</td>
<td>22.0 ± 6.3</td>
</tr>
</tbody>
</table>

*Number of samples; Values are means ± standard deviation; Values with different letter superscripts in the column are significantly different (p < 0.05).*
Table 2. Analysis of fatty acid profiles (% of total fatty acids) in muscles of nine wild freshwater fish species

<table>
<thead>
<tr>
<th>EO*</th>
<th>Fatty acid</th>
<th>C. auratus</th>
<th>C. carpio</th>
<th>P. fulvidraco</th>
<th>A. amblycephala</th>
<th>C. chuatsi</th>
<th>M. meridionalis</th>
<th>A. argus</th>
<th>C. mongolicus</th>
<th>S. asiatica</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>C12:0</td>
<td>0.99 ± 0.06</td>
<td>0.20 ± 0.11</td>
<td>0.16 ± 0.12</td>
<td>0.14 ± 0.06</td>
<td>0.11 ± 0.07</td>
<td>0.14 ± 0.05</td>
<td>0.09 ± 0.07</td>
<td>0.07 ± 0.02</td>
<td>0.05 ± 0.01</td>
</tr>
<tr>
<td>2</td>
<td>C14:0</td>
<td>1.26 ± 0.40</td>
<td>1.87 ± 0.83</td>
<td>2.73 ± 1.52</td>
<td>2.48 ± 1.15</td>
<td>2.06 ± 0.48</td>
<td>1.65 ± 1.12</td>
<td>2.14 ± 1.41</td>
<td>1.87 ± 0.45</td>
<td>1.93 ± 0.17</td>
</tr>
<tr>
<td>3</td>
<td>C15:0</td>
<td>0.71 ± 0.22</td>
<td>0.48 ± 0.20</td>
<td>0.57 ± 0.22</td>
<td>0.58 ± 0.16</td>
<td>0.65 ± 0.13</td>
<td>0.47 ± 0.23</td>
<td>0.55 ± 0.23</td>
<td>0.25 ± 0.13</td>
<td>0.27 ± 0.02</td>
</tr>
<tr>
<td>4</td>
<td>C16:0</td>
<td>20.5 ± 1.18</td>
<td>20.24 ± 2.34</td>
<td>21.87 ± 1.23</td>
<td>20.97 ± 0.74</td>
<td>23.50 ± 1.81</td>
<td>21.48 ± 0.76</td>
<td>24.27 ± 2.44</td>
<td>18.47 ± 1.09</td>
<td>24.71 ± 0.2</td>
</tr>
<tr>
<td>5</td>
<td>C17:0</td>
<td>1.19 ± 0.31</td>
<td>1.15 ± 0.64</td>
<td>0.84 ± 0.51</td>
<td>0.79 ± 0.45</td>
<td>1.19 ± 0.43</td>
<td>0.96 ± 0.28</td>
<td>0.70 ± 0.34</td>
<td>0.21 ± 0.20</td>
<td>0.28 ± 0.02</td>
</tr>
<tr>
<td>6</td>
<td>C18:0</td>
<td>8.65 ± 1.09</td>
<td>7.85 ± 0.88</td>
<td>6.22 ± 1.72</td>
<td>6.87 ± 0.83</td>
<td>8.19 ± 1.01</td>
<td>6.69 ± 3.47</td>
<td>7.97 ± 1.77</td>
<td>5.91 ± 1.58</td>
<td>6.19 ± 0.40</td>
</tr>
<tr>
<td>7</td>
<td>C18:1n7</td>
<td>3.72 ± 1.12</td>
<td>3.83 ± 1.02</td>
<td>2.83 ± 1.52</td>
<td>3.64 ± 0.91</td>
<td>3.99 ± 0.54</td>
<td>5.23 ± 0.81</td>
<td>2.92 ± 0.78</td>
<td>2.77 ± 1.89</td>
<td>1.78 ± 0.18</td>
</tr>
<tr>
<td>8</td>
<td>C18:2n6</td>
<td>2.97 ± 0.24</td>
<td>1.40 ± 0.74</td>
<td>1.43 ± 0.62</td>
<td>0.90 ± 0.13</td>
<td>0.79 ± 0.16</td>
<td>1.12 ± 0.29</td>
<td>1.24 ± 0.40</td>
<td>1.72 ± 0.68</td>
<td>1.35 ± 0.06</td>
</tr>
<tr>
<td>9</td>
<td>C18:3n3</td>
<td>0.59</td>
<td>0.54</td>
<td>0.12</td>
<td>0.34</td>
<td>0.34</td>
<td>0.17</td>
<td>0.21</td>
<td>0.14</td>
<td>0.20</td>
</tr>
<tr>
<td>10</td>
<td>C18:3n6</td>
<td>1.04 ± 0.14</td>
<td>1.00 ± 0.31</td>
<td>0.62 ± 0.54</td>
<td>0.81 ± 0.26</td>
<td>0.89 ± 0.27</td>
<td>0.82 ± 0.21</td>
<td>0.15 ± 0.09</td>
<td>0.27 ± 0.19</td>
<td>0.20 ± 0.02</td>
</tr>
<tr>
<td>11</td>
<td>C20:1n9</td>
<td>1.21 ± 0.37</td>
<td>2.03 ± 0.84</td>
<td>0.84 ± 0.68</td>
<td>2.29 ± 1.14</td>
<td>2.44 ± 1.43</td>
<td>1.21 ± 0.43</td>
<td>1.33 ± 0.99</td>
<td>0.40 ± 0.19</td>
<td>0.20 ± 0.04</td>
</tr>
<tr>
<td>12</td>
<td>C20:2n6</td>
<td>2.17 ± 0.86</td>
<td>1.77 ± 0.71</td>
<td>1.72 ± 0.76</td>
<td>1.59 ± 0.58</td>
<td>2.19 ± 0.36</td>
<td>2.40 ± 1.03</td>
<td>2.42 ± 0.86</td>
<td>0.36 ± 0.13</td>
<td>1.73 ± 0.02</td>
</tr>
<tr>
<td>13</td>
<td>C20:3n9</td>
<td>3.90 ± 2.38</td>
<td>6.69 ± 2.39</td>
<td>6.53 ± 1.31</td>
<td>7.14 ± 2.41</td>
<td>11.25 ± 3.28</td>
<td>11.53 ± 4.63</td>
<td>10.37 ± 3.96</td>
<td>2.14 ± 0.83</td>
<td>3.90 ± 0.71</td>
</tr>
<tr>
<td>14</td>
<td>C20:4n6</td>
<td>45.60 ± 6.53</td>
<td>38.67 ± 2.12</td>
<td>25.74 ± 4.43</td>
<td>33.35 ± 6.33</td>
<td>37.11 ± 4.12</td>
<td>38.20 ± 6.76</td>
<td>34.75 ± 8.66</td>
<td>22.47 ± 2.80</td>
<td>19.43 ± 0.45</td>
</tr>
<tr>
<td>15</td>
<td>C20:5n3</td>
<td>16.55 ± 5.01</td>
<td>12.82 ± 4.32</td>
<td>13.15 ± 3.33</td>
<td>13.32 ± 3.51</td>
<td>18.45 ± 3.90</td>
<td>19.83 ± 5.90</td>
<td>15.99 ± 4.59</td>
<td>4.45 ± 1.00</td>
<td>7.24 ± 0.93</td>
</tr>
<tr>
<td>16</td>
<td>C20:5n6</td>
<td>29.06 ± 5.28</td>
<td>25.44 ± 4.01</td>
<td>12.59 ± 3.71</td>
<td>20.02 ± 7.46</td>
<td>18.67 ± 1.35</td>
<td>18.37 ± 5.39</td>
<td>18.76 ± 7.06</td>
<td>18.02 ± 2.25</td>
<td>12.19 ± 0.69</td>
</tr>
<tr>
<td>17</td>
<td>EPA + DHA</td>
<td>11.93 ± 3.76</td>
<td>9.62 ± 3.68</td>
<td>9.43 ± 1.59</td>
<td>9.72 ± 2.70</td>
<td>14.86 ± 4.06</td>
<td>15.11 ± 5.30</td>
<td>12.30 ± 4.22</td>
<td>2.77 ± 0.90</td>
<td>4.53 ± 0.88</td>
</tr>
<tr>
<td>18</td>
<td>Σn-3 + n-6</td>
<td>0.60 ± 0.27</td>
<td>0.52 ± 0.23</td>
<td>1.14 ± 0.41</td>
<td>0.74 ± 0.28</td>
<td>0.99 ± 0.22</td>
<td>1.16 ± 0.41</td>
<td>1.00 ± 0.55</td>
<td>0.25 ± 0.06</td>
<td>0.59 ± 0.42</td>
</tr>
</tbody>
</table>

* Elution order of fatty acids; nd, not detected; Values are means ±standard deviation

Figure 2. The relationship of total lipid content and SFA, MUFA and PUFA profiles in wild freshwater fish species. Different small letters present different types fatty acids: (a) SFA, (b) MUFA and (c) PUFA, respectively.

Among SFA, the abundant fatty acids were palmitic (C16:0, 18.74 to 24.71%) and stearic (C18:0, 5.91 to 8.65%) acid which were in the range of freshwater fish from Turkish (14.52 to 24.74% and 5.63 to 14.80%, respectively) [27]. The sum of the proportion of the two fatty acids to SFA was above 85.72% of the ratio of minimum found in P. fulvidraco among all the species analyzed, but more in Malaysian freshwater fish [32]. In terms of individual fatty acid in the current study, oleic acid (C18:1n9, 24.38%) was the dominant component in both MUFA and total fatty acids, followed by palmitoleic acid (C16:1, 5.10%). The same authors found the similar levels of oleic and palmitoleic acid (C18:1n9, 23% and C16:1, 4.61%, respectively), however, cis-vaccenic acid (C18:1n7, 0.13%) was not detected in most fish species, and the mean levels determined were significantly lower than those of fish examined in the current study (C18:1n7, 3.41%) [32]. Meanwhile, cis-vaccenic acid in fish was not reported in numerous studies [27,31,33]. This may be attributed to cis-vaccenic acid content low enough to be detected. However, it may also be a consequence of common elution with isomers that were not well resolved by some chromatographic columns [28]. The other individual MUFA was characterized by extremely low composition profiles which were below eicosanoid acid level (C20:1n9, 1.72%) for C. mongolicus. PUFA tested in these wild fish were composed of 13 fatty acids. The
predominant fatty acids were linoleic (C18:2n6), α-linolenic (C18:3n3), docosahexaenoic (DHA, C22:6n3), arachidonic (AA, C20:4n6) and eicosapentaenoic (EPA, C20:5n3) acid. The highest share of these fatty acids was observed in C. auratus (C18:2n6, 14.47%) which was in the range of the wild and cultured C. auratus (11.41% and 17.00%, respectively) [34]. AA constituted 8.99% and 5.90% of the total fatty acids in S. chuatsi and C. argus whereas the values (4.8% and 3.6%) respectively obtained by [34] were significantly different from those determined in our study. The two valuable fatty acids EPA and DHA varied from 2.77% in C. mongolicus to 15.11% in S. meridionalis. The mean value for all species was 10.03%, accounting for 73.6% of the n-3 PUFA that was extremely close to the result (72.8%) reported by [17]. The high ratio was obviously linked to the relatively high DHA composition profiles in n-3 PUFA. The proportions of DHA to total fatty acids extracted from muscle tissue substantially exceeded that of EPA in all of the tested fish species. Typically, the lowest percentage share of EPA was 0.63% obtained from both C. mongolicus and C. asiatica. The current study, to our knowledge, was the first to report the fatty acid profiles of the two species for preliminarily chemical composition data. The results, to some extent, appeared to account for the relatively low AA proportions in C. mongolicus and C. asiatica as a result of decreasing competitive inhibition from AA to eicosanoids caused by low n-3 PUFA levels, especially extremely low EPA and DHA contents [35]. Additional nutritional data and dynamic research of fish species are required.

Nutritional composition indices are important in determining the n-3/n-6 PUFA ratio. All ratios ranging from 0.25 to 1.16 were similar to those of Polish freshwater fish [30] and comparable to those of most Malaysian freshwater fish [32] and those calculated by [34] but lower than those detected in wild fish by [17] which varied from 1.0 to 2.5. This discrepancy was greatly attributed to the dependence on the available food base where the fish were caught and other environmental factors. Some differences in the optimal ratio recommendations for healthy human diets are also reported. Sargent [36] recommended an optimal ratio of 0.2. However, Simopoulos [37] indicated that optimal ratio may vary in consideration of complexities and differences of disease, and the n-3/n-6 ratio range of 0.25 to 1.0 was proposed as a dietary intake standard. Considering the nutritional benefits, The Food and Agriculture Organization and the World Health Organization suggested a high ratio of > 0.2 [38]. Thus, the results from this study were in accordance with the recommended values above and indicate that these fish species are a good food selection for consumption.

3.3. Specific Fatty Acid Quantity in Muscle

The expression of fatty acid quantities in fish is desirable as a reference for human dietary intake. Table 3 showed the fatty acid amounts expressed as mg 100 g⁻¹ wet weight fish muscle. The relatively high amounts of SFA, MUFA, and PUFA were in C. asiatica at 643.0, 899.9 and 369.8 mg 100 g⁻¹ fish muscle tissue, followed by those in P. fulvidraco and C. mongolicus. C. argus had low fatty acid amounts, followed by S. meridionalis and C. auratus. N-3 PUFA content in P. fulvidraco with the highest DHA and EPA concentrations was evidently higher than those in other fish species. In combination with composition profiles, to some extent, it is easily observed that the amounts of fatty acids have a positive relationship with the lipid content of fish [25]. The quantities of DHA and EPA in S. chuatsi and S. meridionalis were 77.8 and 69.1 mg 100 g⁻¹ wet weight, respectively, but lower than that in C. asiatica (84.1 mg 100 g⁻¹) and higher than those in M. amblycephala and C. auratus (64.4 and 55.9 mg 100 g⁻¹, respectively). However, EPA concentration of these wild freshwater fish species fluctuated at an exceedingly low level from 7.7 mg 100 g⁻¹ in C. argus to 19.1 mg 100 g⁻¹ in C. auratus apart from P. fulvidraco. This indicated that EPA from these fish is inferior to DHA as nutritional supplement in human diet.

### Table 3. Fatty acid quantities in muscle of nine wild freshwater fish species *

<table>
<thead>
<tr>
<th>Fatty acids</th>
<th>C. auratus</th>
<th>S. chuatsi</th>
<th>M. amblycephala</th>
<th>C. mongolicus</th>
<th>C. asiatica</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>ΣSFA</strong></td>
<td>164.5 ± 86.0</td>
<td>192.7 ± 54.8</td>
<td>641.3 ± 197.4</td>
<td>210.5 ± 46.5</td>
<td>185.3 ± 78.3</td>
</tr>
<tr>
<td><strong>ΣMUFA</strong></td>
<td>122.0 ± 118.1</td>
<td>177.3 ± 92.1</td>
<td>594.0 ± 259.5</td>
<td>226.0 ± 62.2</td>
<td>138.3 ± 74.6</td>
</tr>
<tr>
<td><strong>ΣPUFA</strong></td>
<td>220.8 ± 90.6</td>
<td>223.4 ± 77.1</td>
<td>358.6 ± 144.9</td>
<td>215.7 ± 55.0</td>
<td>219.9 ± 91.9</td>
</tr>
<tr>
<td>n-6</td>
<td>142.6 ± 69.6</td>
<td>155.7 ± 75.2</td>
<td>177.4 ± 84.8</td>
<td>127.6 ± 41.4</td>
<td>95.3 ± 40.8</td>
</tr>
<tr>
<td>n-3</td>
<td>78.2 ± 43.1</td>
<td>67.6 ± 13.7</td>
<td>181.2 ± 73.5</td>
<td>88.1 ± 33.2</td>
<td>96.6 ± 53.7</td>
</tr>
<tr>
<td>EPA</td>
<td>19.1 ± 12.7</td>
<td>49.9 ± 4.1</td>
<td>40.9 ± 28.3</td>
<td>16.9 ± 4.9</td>
<td>18.3 ± 9.8</td>
</tr>
<tr>
<td>DHA</td>
<td>36.8 ± 18.7</td>
<td>35.0 ± 6.7</td>
<td>87.6 ± 27.0</td>
<td>47.5 ± 22.9</td>
<td>59.5 ± 37.6</td>
</tr>
<tr>
<td>EPA+DHA</td>
<td>55.9 ± 30.1</td>
<td>49.9 ± 10.4</td>
<td>128.4 ± 44.3</td>
<td>64.4 ± 26.6</td>
<td>77.8 ± 47.3</td>
</tr>
</tbody>
</table>

*Results are expressed mg fatty acids per 100 g wet fish muscle; Values are means ± standard deviation.

3.4. Nutritional Quality Assessment

Due to relatively high amounts of EPA and DHA found in fish, investigations and explanatory trials on these compounds associated prevention and treatment of chronic conditions have been conducted for recent decades. On one hand, EPA and DHA intake from fish consumption is negatively related to the onset of diseases such as coronary heart disease (CHD) in numerous studies [6]. Mozaffarian and Rimm [39] recommended daily fatty acids intake of approximately 250 mg for the general population. Compared to wild fish species analyzed in this study with respect to EPA and DHA contents, P. fulvidraco is a better option (Table 3). One 200 g portions of P. fulvidraco daily could meet the requirements for EPA and DHA, and other fish would be asked to more consumption by persons. However, it is unfavorable compared with most marine fish, such as Baltic salmon and herring [30]. But consumption of these wild fish is still recommended to boost health because other animal foodstuffs such as pork, lamb and beef provide little-to-no contribution in this aspect provided that equal quality
intake. On the other hand, IA and IT are widely applied to assess nutritional quality based on fatty acid profiles [40,41,42]. Figure 3 showed that there was a significant difference in both the indices across fish species (p < 0.001). The highest IA and IT values in this study were 0.52 in C. argus and 0.64 in C. asiatica, the lowest value was 0.36 in C. mongolicus and S. meridionalis, respectively. High IA and IT values have adverse effects on human health and are considered to be important factors in underlying CHD risk increase, where the higher the values the more likely the food quality is to be degraded [21]. Increased risk of subsequent Ischemic heart disease is also related to either high IA or IT value [42]. Both of the indices in our study revealed the potential high-quality food for these fish species. The average values of IA and IT from the nine fish species were 0.45 and 0.47, respectively. These values were similar to the mean values of two kinds of the most popularly freshwater fish Hypophthalmichthys nobilis and Ctenopharyngodon idellus in Asia (0.46 and 0.48, respectively) [43]. Meanwhile, the indices were lower than those of seven freshwater fish obtained from the Brazilian Pantanal and were comparable with those of commonly consumed freshwater fish from different countries [25]. Higher IA and IT values in chicken and beef were determined than those from fish in current study [21]. Therefore, fish should not only be considered a popular food because of its culinary properties but also for its health benefits which participate in the prevention and support of medical therapies for chronic conditions. A negative effect sometimes is imposed by accumulative persistent organic pollutants (PCBs and PEDDs) or other toxic chemicals in fish with environmental destruction [44]. Thus, this issue also requires attention, and further study on these subjects is urgent and necessary [45].

![Figure 3. Indices of atherogenicity and thrombogenicity (IA and IT) based on fatty acid compositions of nine wild freshwater fish species](image)

4. Conclusion

Nutritional information regarding fatty acid compositions and nutritional quality for total lipids in nine wild freshwater fish species from China was provided in our study in response to the limited knowledge on this topic. This study revealed that the muscles of all fish species analyzed were characterized by relatively high moisture and low lipid content and significant negative correlations were observed despite differences in species. MUFA and PUFA were positively and negatively correlated with total lipid content, respectively, whereas an indiscernible trend was observed in saturated fatty acids, indicating that a dynamic interaction exists between them what is needed for more attentions in the future. In perspective of nutritional value of these fish species, our results would suggest that these fish are an effective source of essential fatty acids, especially DHA in the diet and reveal that all fish species are a good functional food for the potential health benefits in disease prevention.

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