Evaluation of the Toxicity of Three Hair Shampoos on the Catfish (Clarias gariepinus) Fingerlings

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Abstract The toxicity of three brands of hair shampoos (Vinoz, Gentelle and Petals) containing linear alkylbenzene sulphonates and used mostly for hair conditioning in beauty shops by woman in Nigeria were evaluated on fingerlings of the African catfish, Clarias gariepinus (mean weight=6.42 ± 0.2g; length=4.03 ± 0.1 cm). Specimens were exposed to 0.50%, 0.25%, 0.125%, 0.0625%, 0.03125% and 0.00% (control) toxicant concentrations during a 96-hours static bioassay. Mortality increased with concentrations and varied with brand of shampoo used; with the 0.50% toxicant concentration and Vinoz shampoo showing highest toxicity. LC₅₀ for Vinoz, Gentelle and Petals were 0.0195%, 0.0281% and 0.0281% (arithmetic method), 0.9058%, 0.8873% and 0.8710% (logarithmic method) and 0.0172%, 0.0281% and 0.0177% (probit method) respectively. Survivorship of C. gariepinus was significantly homogeneous with Vinoz [F(1.56)<Fcrit(4.13)], heterogeneous with Gentelle [F(4.31)>Fcrit(4.13)] and heterogeneous with Petals shampoos [F(5.57)>Fcrit(4.13)] at P<0.05. The numbers of survivors in the 0.125 and 0.03125% toxicant concentrations differed significantly with those in the control (Sig. t=0.001 & 0.002 with Vinoz and =0.002 & 0.003 with Gentelle, respectively) at the 95% confidence limit. Very low mean lethal concentrations of the xenobiotics exerted high toxicities on the aquatic biota. Regulatory agencies should enforce strict laws for the protection of the national aquatic biotopes for sustainability.

Keywords: surfactants, toxicity, bioassay, LC₅₀, mortality, xenobiotics


1. Introduction

Surfactants are synthetic organic chemicals used in detergents, personal care and household cleaning. These compounds usually comprise 10%-18% of granular and liquid detergents and are the largest ingredient of the 20-25 compounds used in these products [1]. Hair shampoo is a liquid detergent used mostly by women in conditioning and washing their hair. In Owerri, the Imo State capital and many other Nigerian cities and the hinterlands for example, the establishment of hair dressing saloons is on the increase probably because of modernization and the quest for fashion especially among women. The effluents (wastewater) from these saloons empty into water bodies in their proximity via gutters and urban drains and thus, could pollute them [2] in especially, Sub-Saharan Africa where there are little or no prohibitions for violators.

Beautification is one of man’s body care process that involves the heavy and prolific use of surfactants, which are basically ingredients of body care products. They are known to reduce surface tension in water and allow aqueous solutions to spread and penetrate more easily. Thus, their inclusion in body shampoos enable intended effects of other ingredients to permeate the target body areas such as face, hair, skin etc. Unfortunately, these same characteristics could adversely affect aquatic lives [3], such as altering the properties of a fish’s gills and subsequently change the fish’s normal uptake of ions from the water column [4,5]. Other effects could include tainting of the fish’s body, bioaccumulation of the toxicant in fish tissues or even mortality [6,7]. Unfortunately, the possible toxic effects of these inorganic pollutions have not been investigated in our local environment, against its increasing use and introduction of newer brands by manufacturers. This research therefore was an attempt to close this gap in knowledge, through the evaluation of the toxicity of effluents of three commonly used hair shampoos (Vinoz, Gentelle and Petals) in Owerri Metropolis, southeastern Nigeria, on the popular fish delicacy - Clarias gariepinus.

2. Materials and Methods

An The scientific methods employed in this study were in keeping with standard protocols of Buikema and Cairns [8], Bellan [9], Ward and Parrish [10,11] and Reish and Oshida [12,13].

2.1. Study Area
The Pollution Control Laboratory of the Department of Environmental Technology, Federal University of Technology, Owerri (FUTO), Nigeria where the research was carried out is in Owerri West Local Government Area of Imo State, Southeastern Nigeria. FUTO lies between latitudes 05° 22´ and 05° 39´ and longitudes 06° 08´ and 06° 41´, near Ihiagwa town.

2.2. Acquisition and Acclimation of Fish Specimens

Healthy specimens of the African catfish, *C. gariepinus* of mixed sex and broodstock (mean weight= 6.42 ± 0.2g; mean length= 4.30 ± 0.1cm) were purchased from a fish farm in Owerri. The choice of *C. gariepinus* was made because of its ability to withstand stress and its high commercial value in Nigeria. They were maintained in the laboratory in a large plastic bowl of about 120 litres capacity containing 60 litres of clean borehole water for a minimum period of two weeks for acclimation during which period they were fed 3 times daily (morning, afternoon and night) with commercial fish pellets. Three-quarters of the water was changed every morning at feeding time to reduce constant dirt occasioned by the fish faeces. However, feeding was discontinued 24 hours before commencement of definitive experiment.

2.3. Determination of Physicochemical Properties of Diluent Water

The temperature, Dissolved Oxygen (DO) and pH of experimental water were continuously determined in-situ with a pre-calibrated HANNA HI 9828 PH/ORP/EC/DO meter.

2.4. Active ingredients of Toxicants

Hair-care shampoos essentially contain linear alkybenzene sulphonates and fragrance formulations. The specific chemical compositions of the toxicants used in this study are shown in Table 1.

2.5. Range-finding Test

Ten fish of approximately same sizes were distributed randomly into each of 5 plastic aquaria containing 60 litres of borehole water. Graded concentrations (5, 4, 3, 2 and 1%) of each of the toxicants were serially added to the aquaria and the set-ups allowed to stand for 96 hours. Observations for death of fish were made after 1, 2, 4, 8, 16, 24, 48, 72, and 96 hours. After the 96th hour, only the 1% diluent water-toxicant setup had live fish, while 100% mortalities were recorded in the other toxicant concentration setups.

2.6. Definitive Static Bioassay

Definitive graded concentrations of 0.5, 0.25, 0.125, 0.0625, 0.03125 and 0.00% (control) for each toxicant was constituted in 6 aquaria containing 60 litres of diluents water. Ten fish specimens were added to each of the aquaria. Replicate aquaria were also established and the setup observed for mortalities after 1, 2, 4, 8, 16, 24, 72 and 96 hours. Death was ascertained when fish did not react to gentle poke with a glass rod and was immediately removed to avoid decay and contamination. Loss of equilibrium, vigorous movement of gulping of air and other behavioural patterns were also observed for fish in each plastic aquarium. While the experiment lasted, there were no mortalities in the control aquaria. The test fish were not fed to prevent remnants of uneaten food from contaminating the water thereby increasing mortality rates.

### Table 1. Chemical compositions of the three hair shampoos used as toxicant in 96-hours bioassay

<table>
<thead>
<tr>
<th>Shampoos</th>
<th>Vinoz</th>
<th>Gentelle</th>
<th>Petals</th>
</tr>
</thead>
<tbody>
<tr>
<td>Deionized water, antisol, menthol, soda as, sodium laureth sulphate, eucaly, common salt, methyl salicylate, colourant and fragrance</td>
<td>Deionized water, antisol, sodium laureth sulfate, sodium chloride, cocamidopropyl betaine, parfum, glycol stearate, cocamideMEA, disodium EDTA, citric acid, sodium Hydroxide, triethylen glycol, benzyl alcohol, propylene glycol, sodium benzoate, tetrasodium EDTA, magnesium chloride, magnesium nitrate, methylchloroix othiazolinone, methylisothiazolinone, jasminum officinale, methylparaben and sorbic acid</td>
<td>Alkybenzene sulphate, soda ash(light), sodium hydroxide, methyl paraben, colourant, fragrance, De-ionised water and CMC</td>
<td></td>
</tr>
</tbody>
</table>

2.7. Statistical Analysis

Mean lethal concentrations (LC₅₀) of the toxicants used were calculated using the arithmetic, logarithmic and probit methods. The single factor ANOVA was used to determine variance equality in means of survival rates of fish specimens, while post-hoc structure of group means was detected with means plots at P<0.05. A pairwise comparison of survival rates in the different concentrations of toxicants was conducted with the student’s t-test of significance at the 95% confidence interval.

### 3.1. Mortality of *C. gariepinus* in Vinoz Hair Shampoo

Mortality patterns in replicates 1, 2 and 3 (Figure 1) reveal that 1 fish each survived after 96 hours in the 0.5% and 0.25% toxicant concentrations; in replicates 1 and 2, 1 fish each survived in the 0.125% concentration; and in replicate 3 of the same (0.125%) concentration, 2 fish survived. In the 0.0625% toxicant concentration, 2 fish each survived in replicates 1, 2 and 3 after 96 hour exposure period, whereas in the 0.03125% toxicant concentration, replicate 1 recorded 2 survivors and replicates 2 and 3 recorded 3 survivors each. However, replicates 1, 2 and 3 of the control experiment recorded 100% survivals after the 96 hours. However, a test of homogeneity in mean variance of numbers of survivors among *C. gariepinus* exposed to Vinoz Shampoo revealed
homogeneity [F(1.56)<Fcrit(4.13)] at P<0.05. The mean pH of the experimental setup was 5.2 ± 1.0, temperature was 24.0 ± 0.4°C while DO was 2.1 ± 1.4 mg/L over 96 hours period.

3.2. Mortality of C. gariepinus in Gentelle Hair Shampoo

The mortality trend in replicates 1, 2 and 3 (Figure 2) of the 0.5% and 0.125% toxicant concentrations revealed that only 1 fish survived after 96 hours exposure. Replicate 1 of the 0.125% toxicant concentration recorded 1 survivor while 2 survived in replicates 2 and 3 of the same toxicant concentration. The 0.0625% toxicant concentration recorded 3 survivors each in replicates 1, 2 and 3, while 5 fish survived in replicate 1 of the 0.03125% toxicant concentration. Four fish survived in each of replicates 2 and 3 after 96 hour exposure, while all 10 fish survived in replicates 1, 2 and 3 of the control setup after the 96-hour bioassay. Mean pH, temperature and DO of the toxicant/diluents water mixture was as follows: 5.2 ± 1.0, 26.7 ± 0.10°C, and 3.7 ± 1.3 mg/L over the 96 hours period. There was significant heterogeneity in mean survival of fish species at P<0.05 [F(4.31)>Fcrit(4.13)].

3.3. Mortality of C. gariepinus in Petals Hair Shampoo

In replicates 1, 2 and 3 of the 0.5% toxicant concentration, 1 fish each survived after the 96 hours bioassay (Fig. 3). In the 0.25% concentration, replicate 1 recorded 2 survivors while replicates 2 and 3 recorded 1 survivor each. In replicates 1, 2 and 3 of the 0.125% toxicant concentration, 2 fish each survived, in replicates 1 and 2 of the 0.0625% toxicant concentration, only 4 fish survived in each of them while in replicate 3 of the same toxicant concentration, 3 fish survived. Five fish each survived after 96-hour exposure in replicates 1, 2 and 3 of the 0.03125% toxicant concentration. All fish specimens in the control aquaria also survived after the 96-hours bioassay period. The ANOVA test in mean survivals revealed significant heterogeneity [F(5.57)>Fcrit(4.13)] at P<0.05. In this toxicant/diluents water mixture, mean pH, temperature and DO were 5.9 ± 1.0, 26.4 ± 0.2°C and 3.3 ± 1.3 mg/L over the bioassay period.

3.4. Comparison of Survivals in Treatment and Control Concentrations

Results of the pair-wise comparison in the number of survivors of fingerlings of C. gariepinus exposed to the 0.125% and 0.03125% toxicant concentrations of Vinoz shampoo differed significantly with those of the control (sig.t=0.001 and 0.002 respectively) at P<0.05. Similarly, with Gentelle shampoo toxicant solutions, the number of survivors in the 0.125% and 0.03125% concentrations differed significantly with those of their control solutions (sig. t = 0.002 and 0.003 respectively).

3.5. Mean Lethal Concentrations (LC50) of Toxicants

Using the arithmetic graphical method (AGM), LC50s for Vinoz shampoo were 0.0156% for replicate 1, 0.0209% for replicate 2 and 0.0219% for replicate 3. With the logarithmic graphical method (LGM), LC50s were 0.9141% for replicate 1, 0.9016% for replicate 2 and 0.9016% for replicate 3. However, with the probit graphical method (PGM), LC50s for replicates 1, 2 and 3 were 0.0141%, 0.0188% and 0.0188% respectively. Mean LC50, therefore were 0.0195%AGM, 0.9058%LGM, and 0.0172%PGM.

With the AGM, LC50s for Gentelle shampoo were 0.0313% for replicate 1, 0.0266% for replicate 2 and 0.0266% for replicate 3 and with the LGM, LC50s were 0.8710% for replicate 1, 0.8954% for replicate 2 and 0.8954% for replicate 3. With the PGM, LC50s for replicates 1, 2, and 3 were 0.0259%, 0.0266% and 0.0266% respectively. Mean LC50, were 0.0281%AGM, 0.8873%LGM, and 0.0281%PGM.

With Petals shampoo, LC50s with the AGM was 0.0313% for each of replicates 1, 2 and 3 and with the LGM, it was 0.8710% for each of replicates 1, 2 and 3. However, with the PGM, LC50 values for replicates 1, 2, and 3 were 0.0156%, 0.0188% and 0.0188% respectively. Mean LC50, therefore were 0.0313%AGM, 0.8710%LGM, and 0.0177%PGM.
4. Discussion

Acute toxicity studies have been recognized as the first step in determining the water quality requirements of fish and so reveal toxicant concentrations that cause mortality even at short exposure. They have further been used to derive water quality guidelines for regulatory measures [14]. Biological monitoring using a series of assays having different endpoints could allow a sensitive approach to predict the potential risk of shampoos which is helpful in formulating the ‘safe levels’ of such bioaccumulative chemicals. The observed effects of the hair shampoo effluents on fish in the current research indicates that caution should be exercised in allowing these xenobiotics into the aquatic environment, especially in excessive amounts.

Though the fingerlings were not fed during definitive test periods, the diluents water quality underwent observable degradations, most probably due to the effects of the toxicants and their intermediate bi-products [15,16] as well as physiological wastes from the animals. Effect on fingerlings included immediate hyperactivity of the fish body parts, similar to observations made by Ogundiran et al. [17] on the catfish.

Obviously, survival rates in this study increased with decrease in toxicant concentrations of the three brands of shampoos used, especially with Petals shampoo, in consonance with mortalities increasing with concentrations and varying with brand of shampoo; with the 0.5% toxicant concentration and Vinoz shampoo showing the highest toxicity. LC50s for Vinoz, Gentelle and Petals did not differ markedly and were in the order of magnitude Gentelle>Petals>VinozAGM>Vinoz>GC>PetalsLGQ and Gentelle>Petals>VinozPGM. The observed homogeneity in survivorship of C. gariepinus indicates that dilution did not exert marked effect on the toxicity of the xenobiotic. Conversely, the observed heterogeneity in survivorship of the fish at the different concentrations of Gentelle and Petals shampoos clearly indicate that dilution had effects on the toxicity of the xenobiotics. The contributions to mean variations by the 0.0313% and had effects on the toxicity of the xenobiotic. The very low LC50s recorded in this study indicates that high toxicities were induced by very low concentrations of the xenobiotics. This thus classifies these toxicants as deleterious to aquatic biotopes.

5. Summary and Conclusion

Very low concentrations of the hair care shampoos used as toxicants induced high lethal toxicities in the test organisms. Vinoz shampoo was more toxic to Clarias gariepinus fingerlings than Gentelle and Petal Shampoos. All the Shampoos induced toxic stress in form of behavioural changes in the fish. Mortalities increased with increasing concentrations of the toxicants. The very low LC50s recorded in this study indicates that high toxicities were induced by very low concentrations of the xenobiotics. This thus classifies these toxicants as deleterious to aquatic biotopes.

6. Recommendation

Local regulatory agencies such as the National Environmental Standards and Regulations Enforcement Agency (NESREA), Federal Ministry of Environment (FMEnv) and Federal Environmental Protection Agency (FEPA) should strictly monitor and enforce the relevant laws to protect aquatic lives, especially Clarias gariepinus.